

## Circulating MicroRNAs as Potential Biomarkers for Prostate Cancer

Kasomva K<sup>1,2,3</sup>, Ignacimuthu S<sup>\*1,5</sup>, Paulraj MG<sup>1</sup>, Sen A<sup>2</sup>, Sailo S<sup>3</sup>, Raphael V<sup>4</sup>, Puro K<sup>2</sup> and Ngachan SV<sup>2</sup>

<sup>1</sup>Division of Biotechnology & Molecular Biology, Entomology Research Institute, Loyola College, Chennai, India

<sup>2</sup>Division of Animal health, ICAR Research Complex for NEH Region, Umiam, Meghalaya, India

<sup>3</sup>Department of Urology, North Eastern Indira Gandhi Regional Institute of Health and Medical Sciences, Shillong, Meghalaya, India

<sup>4</sup>Department of Pathology, North Eastern Indira Gandhi Regional Institute of Health and Medical Sciences, Shillong, Meghalaya, India

<sup>5</sup>International Scientific Partnership Program, King Saud University, Saudi Arabia

**\*Corresponding author:** Ignacimuthu S, Division of Biotechnology & Molecular Biology, Entomology Research Institute, Loyola College, Chennai, India and Visiting Professor Programme, International Scientific Partnership Program, King Saud University, Saudi Arabia, Tel: 044 2817 8348, Fax: 044 2817 5566 28, E-mail: eriloyola@hotmail.com

**Citation:** Kasomva K, Ignacimuthu S, Paulraj MG, Sen A, Sailo S, et al. (2016) Circulating MicroRNAs as Potential Biomarkers for Prostate Cancer. SAJ Biotechnol 3: 101

**Article history:** Received: 07 November 2016, Accepted: 27 December 2016, Published: 29 December 2016

### Abstract

Prostate cancer (PCa) is the most dominant cancer among men in America and Europe. Currently used clinical diagnostic markers are ineffective due to low specificity and poor sensitivity. A novel biomarker for diagnosis of PCa is required. A large quantity of microRNAs (miRNAs) is built up of 18-23 nucleotides; they are small non-coding and single-stranded, and are important in post-transcriptional regulation of gene expression by degrading or suppressing target gene mRNAs. MiRNAs are implicated in the pathogenesis of prostate cancer; however, they also act as novel target for the therapeutic intervention. They also show promise as biomarkers. In this review, we discuss the role of circulating miRNAs as potential biomarkers for PCa diagnosis.

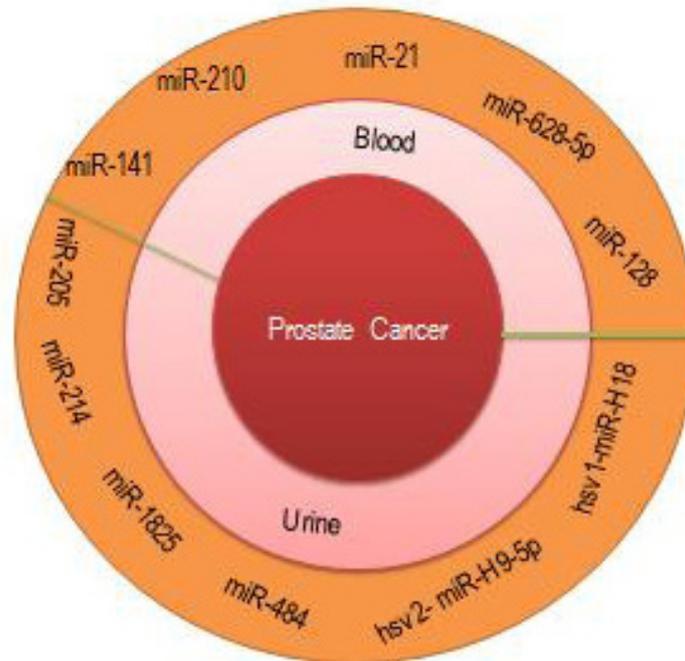
**Keywords:** MicroRNAs (miRNAs); Biomarker; Diagnosis; Prostate Cancer; Post transcriptional

### Introduction

Prostate cancer (PCa) is the most commonly diagnosed malignancy and leading cause of cancer mortality in men worldwide. The American Cancer Society reported that there will be 180,890 new cases of PCa and 26,120 deaths in 2016 [1,2]. PCa development is a slow process commonly in men over the age of fifty years and mainly develops at peripheral zone and involves multiple genetic and molecular changes [3,4]. Prostate specific antigen (PSA) is the only gold standard diagnostic biomarker and indicator for the PCa management. Inclusion of PSA screening into clinical examinations has achieved a significant reduction in PCa-associated death. Although in most of the cases, PSA testing is not disease-specific. As a result, PSA periodical screening led clinicians to misdiagnose and overtreatment the disease and also performed unnecessary prostate biopsies in order to identify a patient with a lethal prostate tumor. This happened due to low specificity and sensitivity as the PSA levels were elevated even in benign prostatic hyperplasia (BPH), chronic inflammation and infection [4-7]. Due to these shortcomings, detection of PCa is not effective. Other biomarkers have been proposed such as total PSA velocity (total PSAV), human glandular kallikrein 2 (hk2), urokinase plasmonogen activator (uPA), urokinase plasmonogen receptor (uPAR), transforming growth factor-beta 1 (TGF- $\beta$  1), interleukin-6 (IL-6) and interleukin-6 receptor (IL-2R) for diagnosis [5,6]. However, developing novel biomarkers to detect with accuracy would greatly assist the diagnosis of PCa.

The importance of microRNAs has been established in different types of cancer including prostate cancer. A large quantity of microRNAs (miRNAs) are built up of 18-23 nucleotides; they are small, non-coding and single-stranded and are important in post-transcriptional regulation of gene expression by degrading or suppressing target gene mRNAs. MiRNAs are active by binding to the 3'-untranslated region of various cancer gene mRNAs. Human encoded genes are composed of 3% of miRNAs and above 30% of mRNAs are regulated by miRNAs which affect various physiological and pathological mechanisms including cell cycle, cell differentiation, cell growth, immune response, metabolism, apoptosis and metastasis. Moreover miRNAs are involved in neuronal, muscle, germline development and embryonic morphogenesis and also participate in stem cell regulation [5,8-10]. Recent studies

have identified miRNAs from various sources such as prostate cell lines, prostate tissue, urine, blood, prostate cancer xenograft, and benign prostatic hyperplasia (BPH) [11,12]. The use of circulating miRNAs as potential non-invasive biomarkers for Pca could become promising (Figure 1).



**Figure 1:** Some microRNAs identified in prostate cancer

## MiRNA Target Detection and Regulation in Prostate Cancer

A large number of miRNAs has been linked to different cellular pathways such as apoptosis, cell proliferation, metastasis and metabolism. About 30% of genes regulation is influenced by miRNAs and 50% of miRNAs are identified within introns of a gene (Table 1) [8,21].

miRNA	Expression	Functions	Role in PCa	Targets	References
miR-34a	Down-regulated	cell proliferation, invasion, promotes anti-apoptotic proteins, metastasis	Tumor suppressor	TCF7, CD44	[13,14]
miR-19a	Up-regulated	Cell proliferation, apoptosis	oncogene	BTG1	[15]
miR-154	Down-regulated	Proliferation, migration, invasion, EMT	Tumor suppressor	CCND2, HMGA2	[16,17]
miR-212	Down-regulated	Autophagy, angiogenesis, senescence, Cell tumor development	Tumor suppressor	SIRT1	[18]
miR-497	Down-regulated	Proliferation, migration, invasion	Tumor suppressor	IKK $\beta$	[19]
MiR-205	Down-regulated	Apoptosis, metastasis	Tumor suppressor	BCL-2	[20]

**Table 1:** Some of the studies on microRNAs in prostate cancer

### MiR-34a

MiRNA-34a is downregulated and functions as tumor suppressor in PCa. Induced MiR-34a expression inhibits cell proliferation, invasion and promotes anti-apoptotic proteins. Ras oncogene is involved in the development and progression of prostate cancer. The effectors are shown to have potential as therapeutic targets for the treatment of androgen-independent prostate cancer. An abnormal WNT/ $\beta$ -catenin signaling is found to be involved in several types of cancers including PCa. The combined activation of Ras and WNT oncogenes leads to the rapid progression of invasive carcinoma from tumorigenesis. MiR-34a is reduced by Ras signaling and functions as a negative regulator of WNT signaling by directly targeting the 3'-UTR of TCF7. Losing of MiR-34a expression leads to an oncogenic effect on cell growth and invasion and in Ras signaling-activated prostate cancer cells [22,23]. In addition, PCa stem cells enhanced clonogenic, tumor-initiating and metastatic capacities; they were enriched in the CD44+ cell population. Enforced expression of MiR-34a in bulk or purified CD44+ prostate cancer cells inhibited clonogenic expansion, tumor regeneration, and metastasis. This revealed that MiR-34a is a key negative regulator of CD44+ prostate cancer cells and showed that CD44 is a direct functional target of MiR-34a to inhibit prostate cancer regeneration and metastasis [24].

### MiR-19a

MicroRNA MiR-19a is upregulated in castration-resistant prostate cancer (CRPC) and functions as oncogene. The inhibition of MiR-19a overexpression in CRPC cells suppressed proliferation and regulated apoptosis. The increased BTG1 expression in CRPC cells also significantly suppressed cell growth and regulated apoptosis. B-cell translocation gene 1 (BTG1) is a family of BTG nuclear protein and it has highest expression in the G0/G1 phases of the cell cycle and is decreased when cells progress through G1 which can inhibit cell proliferation, metastasis, angiogenesis and regulate cell cycle progression and differentiation in various cell types. The study revealed that MiR-19a regulates proliferation and apoptosis of CRPC cells by directly targeting the tumor suppressor gene BTG1 [25-27].

### MiR-154

MicroRNA MiR-154 is downregulated in prostate cancer. Restoration of MiR-154 decreased the potential of prostate cancer cells to grow and proliferate and also down-regulated the expression of Cyclin D2 (CCND2) by binding to 3'-untranslated region. Cyclin Ds (CCNDs) are regulators of cell cycle progression and function as transcriptional co-regulator. The aberrant expression of CCND2 leads to abnormal cells proliferation. Thus, MiR-154 plays an important role in PCa proliferation by suppressing CCND2. In another study MiR-154 was regulating epithelial mesenchymal transition (EMT) by targeting high-mobility group AT-hook 2 (HMGA2). EMT contributes in the process of invasion and metastasis of human cancers and HMGA2 is also involved in the EMT process by aberrant expression in malignant tumors. Forced expressions of MiR-154 or HMGA2 reduced the migration and invasion abilities of PCa cells and inhibited EMT [28,29].

### MiR-212

Autophagy plays a potential role in cancer development and cancer cell survival modulating multiple tumor hallmarks. It serves as a protective process to prevent cancer initiation and promote tumors cell survival and maintenance after neoplastic transformation. Angiogenesis and senescence play vital roles in tumour development. Sirtuin (SIRT) is highly conserved family of lysine modifying nicotinamide adenine dinucleotide (NAD<sup>+</sup>)-dependent class III histone deacetylases; it regulates the signaling pathways by targeting expression of genes involved. SIRT1 is highly expressed in PCa. SIRT1 regulates p300/calcium binding protein, androgen receptor and cell cycle proteins like Rb which are involved in regulating physiological processes such as proliferation, metabolism, differentiation, survival, energy homeostasis, aging and pathological conditions. MiR-212 is down regulated in prostate cancer and plays a vital role in PCa development. Loss of MiR-212 in PCa could lead to induction of autophagy for tumor promoting cell survival and aggressive disease. The negative regulation of MiR-212 in PCa leads to modulation of autophagy, angiogenesis and cellular senescence. SIRT1 modulates autophagy and angiogenesis and also contributes to influencing life span for calorie restriction and senescence in tumor cell growth. Therefore, MiR-212 negatively modulates starvation induced autophagy in PCa cells by targeting SIRT1 and overexpression inhibits angiogenesis and cellular senescence [14,30-32].

### MiR-497

Nuclear factor-kappaB (NF-κB) is a transcriptional factor and plays important roles in various biological processes. Aberrant activation of NF-κB is involved in the progression and also in metastasis and invasion of PCa through matrix metalloproteinase-9 (MMP-9). In a normal pathway, inhibitors of NF-κB (IκBs) kinase β (IKKβ) activate NF-κB by phosphorylation of IκBs. MiR-497, involved in prostate cancer, functions as tumor suppressor and is downregulated in PCa. The aberrant expression of MiR-497 changed cellular proliferation, migration and invasion in PCa cell line PC3-AR by directly targeting IKKβ. The proliferation was suppressed by inducing G0/G1 cell cycle arrest and decreasing of CDK8 protein level after transfection of MiR-497. Thus, MiR-497 repressed the expression of IKKβ and downregulated the activity of CDK8 in PCa [33].

## Urinary MiRNAs as Biomarkers for Prostate Cancer

Urine is one of the most easily accessible and non-invasive biofluids available in urology, nephrology and primary care clinics. The use of a urine RNA-based evaluation appears to be fraught with problems given the known liability of RNA; however, there is increasing evidence that miRNA shows surprising stability in situations in which total RNA or mRNA have been shown to be degraded. Mall *et al.* studied the urinary miRNA stability under various clinically relevant conditions: room temperature and 4 °C, as well as serial freeze-thaw conditions. They revealed that miRNA is relatively stable in the harsh urinary environment; even storing for 5 days at varied temperatures and after ten freeze-thaw cycles and even after trypsin digestion; despite modest degradation, there remained sufficient miRNA for quantitative analysis and did not alter miRNA stability. Further, they suggested that urinary miRNA is best measured within the first 24 h for the most accurate representation of miRNAs in the urine milieu. Moreover, the different miRNAs have shown similar stability when evaluated in plasma and serum [34-36].

MiRNAs that are identified in urine could play an important role as molecular diagnostic markers having non-invasive diagnostic potential. It will overcome the physiological and anatomical problems. In one of the urinary miRNA-based study from 73 urine samples patients with diagnosis of PCa having ≥7 Gleason score and 70 patients diagnosed with BPH revealed that miRNAs are stable molecules which can be measured in urine. This study demonstrated that the elevation of MiR-100 and MiR-200b levels in urine samples was significantly associated with the presence of advanced PCa; moreover MiR-100 was found up-regulated in high-grade prostate intraepithelial neoplasia and also was up-regulated in sera from patients with metastatic castration resistant prostate cancer (mCRPC). Thus, MiR-100/200b could be significant non-invasive biomarker for PCa [4,37,38].

MiR-205 is downregulated and is shown to epigenetically repress tumour suppressor in PCa. It is also reported that MiR-205 is a target of BCL-2 gene, Androgen receptor (AR), and protein kinase C epsilon. The loss of MiR-205 function is associated with low prognosis, apoptosis resistance in PCa and hallmark of epithelial-mesenchymal transition, which lead to metastasis [10,13,33,39-45]. MiR-214 is expressed aberrantly and is downregulated in PCa. Urinary MiR-205 and MiR-214 level together could discriminate PCa patients from healthy individuals with 89% sensitivity and 80% specificity, suggesting that these miRNAs could provide non-invasive molecular biomarkers for detection of PCa [11].

MiR-1825 is upregulated in 88% of PCa patients and has putative targets in member-1 of the Discoidin Domain family of receptors (DDR1). MiR-484 is downregulated in 75% of PCa patients and functions as tumor suppressor; it regulates the expression of E3 ubiquitin-protein ligase (UBR5). MiR-1825 diagnosed PCa by 60% sensitivity and 69% specificity while MiR-484 diagnosed PCa has 80% sensitivity and 10% specificity. The combination of MiR-1825/484 could diagnose PCa by 45% sensitivity and 75% specificity [46,47]. Urine-circulating MiR-21 was significantly in lower levels in PCa patients. In PCa tissues, elevated levels of MiR-21 have been detected which showed significant associations with PCa progression. A higher level of serum MiR-21 was found in both androgen-dependent PCa (ADPC) and hormone-refractory PCa (HRPC) patients with low PSA levels. Moreover, other studies showed the associations between blood MiR-21 levels and aggressive course of PCa. Therefore, the difference of MiR-21 levels in urine and blood can be explained by specific miRNA regulation in separate body compartments and selective secretion of this miRNA into body fluids. MiR-21 plays a crucial role in prostate carcinogenesis by promoting cell proliferation, inhibiting apoptosis, enhancing tumor invasion and metastasis; this miRNA could be considered as a PCa-specific biomarker accessible by lowly invasive ways in urine or blood [48-54]. Therefore, urinary miRNAs could assist in prognosis, diagnosis and prediction of PCa.

## Serum and Plasma MiRNAs as Biomarkers for Prostate Cancer

### MiR-141

Association of circulating miRNAs in PCa had established their possibility to be developed as molecular biomarkers for PCa. Most of the blood based biomarkers are useful for prognosis, diagnosis and effective treatments. One of the pioneering research groups revealed the importance of MiR-141 as circulating biomarker from serum. The authors studied 6 miRNAs from 25 metastatic prostate cancer patients and 25 healthy men. MiR-141 was overexpressed in the PCa when compared to healthy control; MiR-141 showed the greatest differential expression among the other miRNAs. Another study analyzed the expression of 667 miRNAs on serum from advanced PCa patients. In this study MiR-141 and MiR-375 expressions were enhanced in PCa samples and their release into the blood was associated with advanced PCa as compared to the other miRNAs. Therefore, MiR-141 had a sensitivity of 78.9% and specificity of 68.8% in predicting clinical progression [17,34,55,56]. Hence MiR-141 could also be used as a biomarker.

### MiR-210

A study of cancer associated 365 miRNAs which were differentially expressed in serum of metastasis castration resistant PCa and a healthy sample was carried out. Out 365 miRNAs, five serum miRNAs (MiR-141, 200c, 200a, and 375) were increased when compared to healthy 210 samples. The expression of plasma MiR-210 had been reported to be increased in pancreatic patients and was found to be indicator of hypoxia; moreover MiR-210 targeted mTOR in PCa which lead to AKT activation and HIP-1 alpha transcriptional activation. This study revealed that serum MiR-210 levels varied widely amongst mCRPC patients undergoing therapy and showed correlation with the treatment response as assessed by changes in PSA. This indicated that serum MiR-210 can be developed as a predictive molecular biomarker in prostate cancer patients [18,38,57,58].

### MiR-21

MiR-21 is upregulated and functions as onco-microRNA which is negatively modulating the expression of tumor suppressor genes; it is considered as a regulator of androgen receptor. 30 radical prostatectomy (RP) patients (14 patients with rapid biochemical failure (BF) and 16 patients without BF) with Gleason score 7 were analyzed for 1435 miRNAs. MiR-21 expression was significantly upregulated in patients with BF as compared to non-BF group ( $p = 0.05$ ). The stromal expressions of MiR-21 had predictive impact on biochemical failure-free survival (BFFS) and clinical failure-free survival (CFFS). Elevation of MiR-21 expression was an independent prognostic factor for BFFS patients with Gleason score 6. For this type of patients MiR-21 may help to predict the risk of future disease progression [59-64]. In another study 56 patient serum samples (20 localized PCa, 20 androgen-dependent PCa, 10 hormone-refractory PCa administered docetaxel-based chemotherapy and 6 BPH) were tested for MiR-21 expression. It revealed that the expression of serum MiR-21 was correlated to serum PSA level in patients with ADPC and HRPC ( $P=0.012$  and  $0.049$ ) whereas there was no significant difference in serum MiR-21 level between BPH, localized CaP and ADPC. The serum MiR-21 was elevated in HRPC patients, who were resistant to docetaxel-based chemotherapy. Thus this miRNA could be used as a marker to indicate the transformation to hormone refractory disease, and a potential predictor for the efficacy of docetaxel-based chemotherapy [51].

The miRNA profiling of serum samples from 36 African (24 PCa patients and 12 controls) and 36 caucasian American (16 PCa patients and 20 control) patients affected by PCa was studied. Three miRNAs namely MiR-25 ( $p < 0.01$ ), MiR-101 ( $p < 0.001$ ) and MiR-628-5p ( $p < 0.0001$ ) showed low expression in serum of PCa patients as compared to normal. Among all three miRNAs, MiR-628-5p was significantly downregulated in all PCa patients. This revealed that MiR-628-5p could be developed as a non-invasive biomarker for PCa diagnosis and prognosis [65].

## MiR-128

MiR-128 functions as tumor suppressor in neuroblastoma, glioma and PCa and is regarded as a negative regulator of malignant phenotype of PCa such as proliferation, cell motility, invasion, apoptosis, and self-renewal. MiR-128 was significantly decreased in both serum and tissue of PCa patients compared to normal samples; this was associated with aggressive clinical pathological features like advanced pathological stage, positive lymph node metastasis, high preoperative PSA and positive angiolymphatic invasion. Moreover, MiR-128 expression was an independent prognostic factor for biochemical recurrence (BCR) free survival of PCa patients. Therefore MiR-128 can be developed as a noninvasive biomarker for PCa prognosis and diagnosis [15,19,66-68].

Recent study from 149 PCa patients, 57 healthy controls, and 121 BPH and other urinary diseases also showed this.

## Virus MicroRNAs as Biomarker for Prostate Cancer

Many epidemiological studies reported the connection between herpes virus infection and prostate cancer risk as this virus inhibited cell apoptosis and stimulated DNA synthesis. Virus miRNAs are used to control the expression of either the host's genes and/ or their own. The first virus miRNAs were found in cells infected with EBV. About 95 % of virus miRNAs known today are of herpes virus origin. In one of the recent studies of 1052 urine, 150 serum, and 150 prostate tissue samples from PCa or BPH patients overexpression of herpes virus miRNAs in urine samples from prostate cancer patients was detected compared to control subjects. This study revealed that hsv1-MiR-H18 and hsv2- MiR-H9-5p detected in urine samples had better diagnostic biomarker performance than tPSA levels in prostate cancer patients [16,69-73].

## Conclusion

In prostate cancer diagnosis biomarker development is of urgent need. Many researchers have shown the potential role of microRNAs as biomarker in prostate cancer. A number of significant dysregulated microRNAs and aberrant expression of micro RNAs have been reported which are closely related with progression of malignancies of prostate cancer. Specific identification or detection of microRNAs in serum, plasma and urine of prostate cancer patients has been done using various techniques. These developments indicate that circulating microRNAs have potential as biomarkers in prostate cancer.

## Acknowledgment

The authors are grateful to Entomology Research Institute, Loyola College, Chennai, for financial assistance. We also thank ICAR-Research complex for NEH Region, Umiam, Meghalaya and North Eastern Indira Gandhi Regional Institute of Health & Medical Sciences, Shillong, Meghalaya for support in manuscript preparation. We are indebted to Miss Thotyachan Ch from D.M. College of Science, Imphal for helping in collecting research papers.

## References

1. Siegel LR, Miller KD, Jemal A (2016) Cancer statistics, 2016. *CA Cancer J Clin* 66: 7-30.
2. American Cancer Society (2016) Cancer Facts and Figures 2016, USA.
3. Carvalhal GE, Griffin CR, Kan D, Loeb S, Catalona WJ (2010) Reducing blood loss in open radical retropubic prostatectomy with prophylactic periprostatic sutures. *BJU Int* 105: 1650-3.
4. Salido-guadarrama AI, Morales-montor JG, Rangel-escareño C, Langley E, Peralta-zaragoza O, et al. (2016) Urinary microRNA-based signature improves accuracy of detection of clinically relevant prostate cancer within the prostate-specific antigen grey zone. *Mol Med Rep* 13: 4549-60.
5. Srivastava A, Suy S, Collins SP, Kumar D (2011) Circulating MicroRNA as Biomarkers: An Update in Prostate Cancer. *Mol Cell Pharmacol* 3: 115-24.
6. Shariat SF, Semjonow A, Lilja H, Savage C, Vickers AJ, et al. (2011) Tumor markers in prostate cancer I: Blood-based markers. *Acta Oncol* 50: 61-75.
7. Wang J, Ye H, Zhang D, Hu Y, Yu X, et al. (2016) MicroRNA 410-5p as a potential serum biomarker for the diagnosis of prostate cancer. *Cancer Cell Int* 16: 12.
8. Kasomva K (2015) MicroRNA—a small non-coding RNA: An efficient biomarker for prostate cancer.
9. Mahn R, Heukamp LC, Rogenhofer S, Ruecker AV, Muller SC, et al. (2011) Circulating microRNAs (miRNAs) in serum of patients with prostate cancer. *Urology* 77: 10.1016/j.urology.2011.01.020.
10. Brase JC, Wuttig D, Kuner R, Sultmann H (2010) Serum microRNAs as non-invasive biomarkers for cancer. *Mol Cancer* 9: 306.
11. Srivastava A, Goldberger H, Dimtchev A, Ramalinga M, Chijioko J, et al. (2013) MicroRNA Profiling in Prostate Cancer - The Diagnostic Potential of Urinary miR-205 and miR-214. *PLoS ONE* 8: e76994.
12. Liu DF, Wu JT, Wang JM, Liu QZ, Gao ZL, et al. (2012) MicroRNA Expression Profile Analysis Reveals Diagnostic Biomarker for Human Prostate Cancer. *Asian Pacific J Cancer Prev* 13: 3313-7.
13. Gandellini P, Folini M, Longoni N, Pennati M, Binda M, et al. (2009) miR-205 Exerts tumor-suppressive functions in human prostate through down-regulation of protein kinase Cepsilon. *Cancer Res* 69: 2287-95.
14. Green AA, Silver PA, Collins JJ, Yin P (2014) Toehold switches: de-novo-designed regulators of gene expression. *Cell* 159: 925-39.
15. Guidi M, Muiños-Gimeno M, Kagerbauer B, Marti E, Estivill X, et al. (2010) Overexpression of miR-128 specifically inhibits the truncated isoform of NTRK3 and upregulates BCL2 in SH-SY5Y neuroblastoma cells. *BMC Mol Biol* 11: 95.
16. Heidegger I, Borena W, Pichler R (2015) The Role of Human Papilloma Virus in Urological Malignancies. *Anticancer Res* 35: 2513-9.
17. Hessvik NP, Sandvig K, Llorent A (2013) Exosomal miRNAs as biomarkers for prostate cancer. *Front Genet* 4: 36.

18. Ho AS, Huang X, Cao H, Christman-Skieller C, Bennewith K, et al. (2010) Circulating miR-210 as a Novel Hypoxia Marker in Pancreatic Cancer. *Transl Oncol* 3: 109-13.
19. Khan AP, Poisson LM, Bhat VB, Fermin D, Zhao R, et al. (2010) Quantitative proteomic profiling of prostate cancer reveals a role for miR-128 in prostate cancer. *Mol Cell Proteomics* 9: 298-312.
20. Khanmi K, Ignacimuthu S, Pualraj MG (2015) MicroRNA in Prostate Cancer. *Clinica Chemica Acta* 451: 154-60.
21. Mohammad Iffat Kabir Anindo, Yaqinuddin A (2012) Insights into the potential use of microRNAs as biomarker in cancer. *Int J of Surgery* 10: 443-49.
22. Chen WY, Liu SY, Chang YS, Yin JJ, Yeh HL, et al. (2015) MicroRNA-34a regulates WNT/TCF7 signaling and inhibits bone metastasis in Ras-activated prostate cancer. *Oncotarget* 6: 441-57.
23. Mulholland DJ, Kobayashi N, Ruscetti M, Zhi A, Tran LM, et al (2012) Pten loss and RAS/MAPK activation cooperate to promote EMT and metastasis initiated from prostate cancer stem/progenitor cells. *Cancer Res* 72: 1878-89.
24. Lui C, Kelnar K, Liu B, Chen X, Calhoun-Davis T, et al (2011) The microRNA miR-43a inhibits prostate cancer stem cells and metastasis by directly repressing CD44. *Nat Med* 17: 211-5.
25. Lu K, Liu C, Tao T, Zhang X, Zhang L, et al. (2015) MicroRNA-19a regulates proliferation and apoptosis of castration-resistant prostate cancer cells by targeting BTG1. *FEBS Lett* 589: 1485-90.
26. Zhao Y, Gou WF, Chen S, Takano Y, Xiu YL, et al. (2013) BTG1 Expression Correlates with the Pathogenesis and Progression of Ovarian Carcinomas. *Int J Mol Sci* 14: 19670-80.
27. Winkler GS (2010) The mammalian anti-proliferative BTG/Tob protein family. *J Cell Physiol* 222: 66-72.
28. Zhu C, Shao P, Bao M, Li P, Zhou H, et al. (2014) MiR-154 inhibits prostate cancer cell proliferation by targeting CCND2. *Urol Oncol* 32: 31.e9-16.
29. Zhu C, Li J, Cheng G, Zhou H, Tao L, et al. (2013) MiR-154 inhibits EMT by targeting HMGA2 in prostate cancer cells. *Mol Cell Biochem* 379: 69-75.
30. Ramalinga M, Roy A, Srivastava A, Bhattarai A, Harish V, et al. (2015) MicroRNA-212 negatively regulates starvation induced autophagy in prostate cancer cells by inhibiting SIRT1 and is a modulator of angiogenesis and cellular senescence. *Oncotarget* 6: 34446-57.
31. Roth M, Chen WY (2014) Sorting out functions of sirtuins in cancer. *Oncogene* 33: 1609-20.
32. Guo JY, Xia B, White E (2013) Autophagy-mediated tumor promotion. *Cell* 155: 1216-9.
33. Kong XJ, Duan LJ, Qian XQ, Xu D, Liu HL, et al. (2015) Tumor-suppressive microRNA-targets IKK $\beta$  to regulate FN-kB signaling pathway in human prostate cancer cells. *Am J Cancer Res* 5: 1795-804.
34. Mitchell PS, Parkin RK, Kroh EM, Fritz BR, Wyman SK, et al. (2008) Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 105: 10513-8.
35. Schwarzenbach H, Hoon DS, Pantel K (2011) Cell-free nucleic acids as biomarkers in cancer patients. *Nat Rev Cancer* 11: 426-37.
36. Christine M, David MR, Blythe DJ, Robert HW (2013) Stability of miRNA in human urine supports its biomarker potential. *Biomark Med* 7: 10.2217/bmm.13.44.
37. Leite KR, Tomiyama A, Reis ST, Sousa-Canavez JM, Sañudo A, et al. (2013) MicroRNA expression profiles in the progression of prostate cancer - from high-grade prostate intraepithelial neoplasia to metastasis. *Urol Oncol* 31: 796-801.
38. Cheng HH, Mitchell PS, Kroh EM, Dowell AE, Chéry L, et al. (2013) Circulating microRNA Profiling Identifies a Subset of Metastatic Prostate Cancer 307 Patients with Evidence of Cancer-Associated Hypoxia. *Plos One* 8: 10.1371/journal.pone.0069239.
39. Lin PC, Chiu YL, Banerjee S, Park K, Mosquera JM, et al. (2013) Epigenetic Repression of miR-31 Disrupts Androgen Receptor Homeostasis and Contributes to Prostate Cancer Progression. *Cancer Res* 73: 1232-44.
40. Schaefer A, Jung M, Mollenkopf HJ, Wagner I, Stephan C, et al. (2010) Diagnostic and prognostic implications of microRNA profiling in prostate carcinoma. *Int J Cancer* 126: 1166-76.
41. Hagman Z, Hafliadottir BS, Ceder JA, Larne O, Bjartell A, et al. (2013) MiR-205 negatively regulates the androgen receptor and is associated with adverse outcome of prostate cancer patients. *Br J Cancer* 108: 1668-76.
42. Tucci P, Agostini M, Grespi F, Markert EK, Terrinoni A, et al. (2012) Loss of p63 and its microRNA-205 target results in enhanced cell migration and metastasis in prostate cancer. *Proc Natl Acad Sci U S A* 109: 15312-7.
43. Puhr M, Hoefler J, Schafer G, Erb HH, Oh SJ, et al. (2012) Epithelial-to-Mesenchymal Transition Leads to Docetaxel Resistance in Prostate Cancer and Is Mediated by Reduced Expression of miR-200c and miR-205. *Am J Pathol* 181: 2188-201.
44. Hulf T, Sibbritt T, Wiklund ED, Patterson K, Song JZ, et al. (2013) Epigenetic induced repression of microRNA-205 is associated with MED1 activation and a poorer prognosis in localized prostate cancer. *Oncogene* 32: 2891-9.
45. Bhatnagar N, Li X, Padi S KR, Zhang Q, Tang MS, et al. (2010) Downregulation of miR-205 and miR-31 confers resistance to chemotherapy induced apoptosis in prostate cancer cells. *Cell Death Dis* 1: 10.1038/cddis.2010.85.
46. Yang SH, Baek HA, Lee HJ, Park HS, Jang KY, et al. (2010) Discoidin domain receptor 1 is associated with poor prognosis of non-small cell lung carcinomas. *Oncol Rep* 24: 311-9.
47. Haj-Ahmad TA, Abdalla MA, Haj-Ahmad Y (2014) Potential Urinary miRNA Biomarker Candidates for the Accurate Detection of Prostate Cancer among Benign Prostatic Hyperplasia Patients. *J Cancer* 5: 182-91.
48. Stuopelytė K, Daniūnaitė K, Jankevičius F, Jarmalaitė S (2016) Detection of miRNAs in urine of prostate cancer patients. *Medicina* 52: 116-24.
49. Ribas J, Lupold SE (2010) The transcriptional regulation of miR-21, its multiple 421 transcripts, and their implication in prostate cancer. *Cell Cycle* 9: 923-9.
50. Li T, Li RS, Li YH, Zhong S, Chen YY, et al. (2012) miR-21 as an independent biochemical recurrence predictor and potential therapeutic target for prostate cancer. *J Urol* 187: 1466-72.

51. Zhang HL, Yang LE, Zhu Y, Yao XD, Zhang SL, et al. (2011) Serum miRNA-21: elevated levels in patients with metastatic hormone-refractory prostate cancer and potential predictive factor for the efficacy of docetaxel-based chemotherapy. *Prostate* 71: 326-31.
52. Yaman Agaoglu F, Kovancilar M, Dizdar Y, Darendeliler E, Holdenrieder S, et al. (2011) Investigation of miR-21, miR-141, and miR-221 in blood circulation of patients with prostate cancer. *Tumour Biol* 32: 583-8.
53. Shen J, Hruby GW, McKiernan JM, Gurvich I, Lipsky MJ, et al. (2012) Dysregulation of circulating microRNAs and prediction of aggressive prostate cancer. *Prostate* 72: 1469-77.
54. Krichevsky AM, Gabriely G (2009) miR-21: a small multi-faceted RNA. *J Cell Mol Med* 13: 39-53.
55. Brase JC, Johannes M, Schlomm T, Fälth M, Haese A, et al. (2011) Circulating miRNAs are correlated with tumour progression in prostate cancer. *Int J Cancer* 128: 608-16.
56. Nikhil S, Luke AS (2013) Circulating MicroRNAs as Biomarkers of Prostate cancer: The state of play. *Prostate cancer* 2013: 10.1155/2013/539680.
57. Wang J, Chen J, Chang P, LeBlanc A, Li D, et al. (2009) MicroRNAs in plasma of pancreatic ductal adenocarcinoma patients as novel blood-based biomarkers of disease. *Cancer Prev Res* 2: 807-13.
58. Majumder PK, Febbo PG, Bikoff R, Berger R, Xue Q, et al. (2004) mTOR inhibition reverses Akt-dependent prostate intraepithelial neoplasia through regulation of apoptotic and HIF-1-dependent pathways. *Nat Med* 10: 594-601.
59. Hulf T, Sibbritt T, Wiklund ED, Bert S, Strbenac D, et al. (2011) Discovery pipeline for epigenetically deregulated miRNAs in cancer: integration of primary miRNA transcription. *BMC Genomics* 12: 10.1186/1471-2164-12-54.
60. Ribas J, Ni X, Haffner M, Wentzel EA, Salmasi AH, et al. (2009) MiR-21: an androgen receptor-regulated microRNA that promotes hormone-dependent and hormone-independent prostate cancer growth. *Cancer Res* 69: 7165-9.
61. Folini M, Gandellini P, Longoni N, Profuma V, Callari M, et al. (2010) MiR-21: an oncomir on strike in prostate cancer. *Mol Cancer* 9: 12.
62. Coppola V, De Maria R, Bonci D (2010) MicroRNAs and prostate cancer. *Endocr Relat Cancer* 17: F1-17.
63. Li T, Li D, Sha J, Sun P, Huang Y (2009) MicroRNA-21 directly targets MARCKS and promotes apoptosis resistance and invasion in prostate cancer cells. *Biochem Biophys Res Commun* 383: 280-5.
64. Melbø-Jørgensen C, Ness N, Andersen S, Valkov A, Dønnem T, et al. (2014) Stromal Expression of MiR-21 Predicts Biochemical Failure in Prostate Cancer Patients with Gleason Score 6. *Plos One* 9: 10.1371/journal.pone.0113039.
65. Srivastava A, Goldberger H, Dimtchev A, Marian C, Soldin O, et al. (2014) Circulatory miR-628-5p is downregulated in prostate cancer patients. *Tumour Biol* 35: 4867-73.
66. Zhang Y, Chao T, Li R, Liu W, Chen Y, et al. (2009) MicroRNA-128 inhibits glioma cells proliferation by targeting transcription factor E2F3a. *J Mol Med* 87: 43-51.
67. Medina-Villaamil V, Martínez-Breijo S, Portela-Pereira P, Quindós-Varela M, Santamarina-Caínzos I, et al. (2014) Circulating microRNAs in blood of patients with prostate cancer. *Actas Urol Esp* 38: 633-9.
68. Sun X, Yang Z, Zhang Y, He J, Wang F, et al. (2015) Prognostic implications of tissue and serum levels of microRNA-128 in human prostate cancer. *Int J Clin Exp Pathol* 8: 8394-401.
69. Ge X, Wang X, Shen P (2013) Herpes simplex virus type 2 or human herpesvirus 8 infection and prostate cancer risk: A meta-analysis. *Biomed Rep* 1: 433-9.
70. Caini S, Gandini S, Dudas M, Bremer V, Severi E, et al. (2014) Sexually transmitted infections and prostate cancer risk: a systematic review and meta-analysis. *Cancer Epidemiol* 38: 329-38.
71. Qureshi A, Thakur N, Monga I, Thakur A, Kumar M (2014) VIRmiRNA: a comprehensive resource for experimentally validated viral miRNAs and their targets. *Database* 1-10.
72. Yun SJ, Jeong P, Kang HW, Kim YH, Kim EA, et al. (2015) Urinary MicroRNAs of prostate cancer: virus-482 encoded hsv1-miRH18 and hsv2-miR-H9-5p could Be valuable diagnostic markers. *Int Neurourol J* 19: 74-84.
73. Kim J, Yun SJ, Kim WJ (2015) Virus encoded circulatory miRNAs for early detection of prostate cancer. *BMC Urol* 15: 116.