

RESEARCH ARTICLE

A Survey of Some Varieties of *Canarium schweinfurthii* (Atili) Grown in Some Parts of Jos East LGA, Plateau State, Nigeria and their Antibacterial Activities

Nyam MA^{1*}, Obashola OE² and Dawang S³

¹Department of Plant Science and Technology, University of Jos, Plateau State, Central Nigeria

²Federal College of Forestry, Jos, Plateau State, Central Nigeria

³Department of Plant Science and Technology, University of Jos, Plateau State, Central Nigeria

*Corresponding author: Nyam MA, Department of Plant Science and Technology, University of Jos, Plateau State, Central Nigeria, Tel: +2348037196083, E-mail: drnyamagm@gmail.com

Citation: Nyam MA, Obashola OE, Dawang S (2018) A Survey of Some Varieties of *Canarium schweinfurthii* (Atili) Grown in Some Parts of Jos East LGA, Plateau State, Nigeria and their Antibacterial Activities. J Microbiol Lab Sci 1: 102

Abstract

A survey was undertaken in Fobur district of Jos-East Local Government Area of Plateau State to document the varieties of *Canarium schweinfurthii* and their antimicrobial activities. Three (3) villages were selected and five (5) questionnaires were administered in each village. Information on the fruit ripening, flowering periods, accessory attributes, harvesting were ascertained. Also information on weight, length, width, shapes and pre-warming duration were determined. The biochemical determinations and antimicrobial activities of leaves extract on *Pseudomonas aeruginosa*, *Salmonella typhi*, *Staphylococcus aureus*, *Streptococcus mutans*, *Bacillus subtilis* and *Escherichia coli* were investigated. The phytochemical screening was carried out using a standard method while agar well diffusion method was adopted for the antimicrobial activity. The biochemical analysis of leaves extract showed the presence of flavonoids, steroids, tannins, saponins, terpenoids, carbohydrates and anthraquinones. Alkaloids and cardiac glycosides were absent. Also the phytochemical screening of fruits revealed the presence of carbohydrates, flavonoids and steroids only. The leaf extract from Kerker village inhibited all test organism at all concentrations (400 mg/ml, 200 mg/ml, 100 mg/ml and 50 mg/ml) except for *Streptococcus mutans* the average fruit length was 4.3 cm, width, 7.5 cm with an ovate shape taking 8 minutes to prewarm. The leaf extracts from Rizek village inhibited the test organisms except *Pseudomonas aeruginosa*, the average fruit length was 3.9cm width 6.1cm also an ovate shape, taking 11 minutes to prewarm. While the leaf extracts from Nubatong village only inhibited *Streptococcus mutans*, *Bacillus subtilis* and *Escherichia coli*. The fruits average length was 4.3 cm, width 6.3 cm and the pre-warming duration of 12 minutes. The reference drug (ciprofloxacin) gave a significant inhibition zone as compared with test extracts. The study has revealed the different varieties of fruit based on shapes, flowering period, ripening period and other attributes which have been documented. The Minimum Inhibitory Concentration (MIC) on the test organisms was 25 mg/ml and the Minimum Bactericidal Concentration on test organism was 12.5 mg/ml. Based on the findings from this study, *Canarium schweinfurthii* leaf have revealed active bacterial tendency against microorganisms used.

Keywords: *Canarium schweinfurthii*; Antimicrobial; Phytochemicals; Leaf Extracts; Jos

Introduction

In the last few decades, there have been growing concerns and realization among experts that fruits should no longer be considered a luxury but a necessity, since they are essentially good for maintenance of health [1]. Fruits and seeds of plants are a good source of food for humans, including many healthy fats, such as omega fats. According to Gordon (1999), experts recommended the consumption of at least 5.7 g of fruits in our daily diet in addition to cereal, pulse, milk and vegetables.

The survey of a country's natural resource is an important prerequisite for proper utilization of the raw materials; it also explores the use of plants for shelter, food, medicine, clothing and religious ceremonies [2,3].

Canarium schweinfurthii is one of those plants usually used by indigenous people for its nutritional value; it belongs to the family *Burseraceae*. A perennial plant found in Africa referred to as black olive, African olive, black elemi etc [4]. In English, it is

commonly known as purple canary tree, bush candle amongst others. The tree has geographical distribution throughout Africa, native species found in Angola, Cameroun, Ethiopia, Ghana, Guinea-Bissau, Liberia, Mali, Senegal, Sierra-Leone, Sudan, Tanzania, Togo, Uganda, Zambia and Nigeria [5]. In Nigeria, Nyam, reported that the tree is common in some parts of states like Bauchi, Plateau, Niger, Southern Kaduna and Oyo [6]. In Plateau State, it is found in Pankshin, Bokkos, Jos North, Jos South and Jos East. According to Burkill, the tree grows up to 40m high or more. The fruit is similar in structure and colour to the well-known fruit of the olive [7,8]. The flowers grow in clusters at the end of the twigs, small and green in colour [9]. The fruit has a hard stone seed that is edible and oily [10]. According to Dawang, *et al.*, the tree is a major source of oleoresin that is used in food medicine and has a range of industrial application, [11,12]. Usually presented by local people. The pulp contains 71 % palmitic acid and 18 % oleic acid which is edible Orwa, *et al.*, [5].

The use of plants and plant parts in curing human ailments has been documented in various ancient manuscript such as Bible, Koran, Rigvedas and Material medical of the ancient Greek philosopher Discorides [13]. In Native American, 1,625 species of plants have been used as food, over 2,500 have been found useful for drugs [14,15]. He also reported that approximately 18,000 species of plants are neither use for drugs nor food. There are very ancient references for utilization of plants in clinical treatments, this information of using plants part for clinical treatment is very well known to common people living in rural areas of developing countries [16].

Today, medicinal plants are generally embraced by most population in the world especially in the developing countries because they have been discovered to hide the best medicine that nature can offer to mankind. They are also cheaper and more accessible than the conventional orthodox medicines [17,18]. Lewis, *et al.* reported that mainstream medicine was increasingly receptive to the use of antimicrobial and other drugs that are plant based as traditional antibiotics become ineffective, as new particularly viral diseases remain intractable to this type of drug [19,20]. Another driving factor for the renewed interest in plant antimicrobials in the past 20 years has been rapid rate of plant species extraction.

Uses of *Canarium schweinfurthii*

The edible fruits can be consumed when softened in warm water to boost the taste. It can serve as an ingredient for preparing dishes. It can be cooked and processed into fruity butter that can serve as a shear butter substitute [21]. The resin collected from the tree is prepared into herbs that treat and fight against intestinal worms such as roundworm [22]. The tree bark is purgative and can be decocted and used for treating gastrointestinal diseases. The leaves can be squeezed and used alone or can be combined with other herbs as for treating cold and cough. The oleoresin from the tree is an important resin used for fumigating and making body creams and paints. The residue from the combustion of the resins is gathered as carbon black that can be used for producing ink. The seeds can be used for ornamental purposes such as making necklaces, bangles and costumes. The seeds can also be used for making local instruments. The wood is used for constructing boats, mortars, flooring and furniture [21]. The mineral composition of the pulpy fruit has been reported to contain 5.6 % protein, 30-35 %, fats, 8.2 % starch, 11.8 cellulose 8.3 % ash, Potassium 1.2 % and calcium 0.4 % [23].

Materials and Methods

Jos East Local Government Area of Plateau State has its headquarters at Angware with an area of 1,020km² and population of 85,602 (Census, 2006).

SAMPLING TECHNIQUE

Jos East has 5 districts, purposive sampling was employed to select Fobur district and three villages (Kerker, Nubatong, Rizek) were selected using simple random sampling technique.

Method of Data Collection: A structured questionnaire 'tiled A survey of varieties of *Canarium schweinfurthii* grown in Fobur district of Jos East LGA of Plateau State was designed to document the varieties. Five (5) questionnaires were purposively administered in each village

Methods

Collection of Plant Sample: Leaves and fruits of *Canarium schweinfurthii* were collected at the study site in January, 2017.

Sample Preparation: The plants sample (leaves) were washed and air-dried at 37 °C for two weeks. The dried samples were then pulverized into powdered form using mortar and pestle for easy extraction of the plant extract. The fruits were also air-dried and removed from the seed with the aid of a knife and blended into fine powdered form.

Ethanol Extraction of Plant material: About 50 g of the powdered leave were weighed into 500ml conical flask and was soaked in ethanol. These were left to stand overnight (24 hours) and shake on a mechanical shaker for 3 hours. The content was filtered using a non-absorbent cotton wool on Buchner funnel-flasks using a vacuum pump. The residue was subjected to several parts of rinsing and filtration with fresh solvents to attain some levels of exhaustive extraction. The collective filtrates were evaporated to dryness using a rotary evaporator. The percentage yield of the extracts were determined as the extracts transferred into a stirrer sample container and preserved in the refrigerator.

Phytochemical Screening of the Plant Extract

The phytochemical screening of the plants extracts (leaves and fruits) were carried out using standard qualitative methods [24,25]. They were tested for alkaloids, tannins, saponins, flavonoids, carbohydrates, cardiac glycosides, steroids, anthraquinones and terpenoids.

Antimicrobial Screening

Sources of Microorganisms

Pure isolates of gram positive and gram negative *Escherichia coli*, *Bacillus subtilis*, *Pseudomonas aeruginosa*, *Salmonella typhi*, *Streptococcus mutans* and *Staphylococcus aureus* were obtained from the pharmaceutical Microbiology Department, University of Jos.

Preparation of the Isolates

A sterile wire loop was used to pick colonies of the organism from the fresh culture prepared by sub culturing. This was inoculated in six bijoux bottles containing 5ml sterile nutrient broth. The organisms were inoculated at 37 °C for 24 hours.

Preparation of Different Concentration of Extracts

The different crude extracts were reconstituted with sterile distilled water to obtain stock solution. 4.0 g of extract were dissolved in 10 ml sterile water to obtain a concentration of 400 mg/ml of plant extract. The stock was diluted serially into 200 mg/ml, 100 mg/ml and 50 mg/ml. A control (ciprofloxacin 500 mg) was dissolved in 10 ml distilled water making it 50mg, it was further diluted with 10 ml sterile water to make it 5 mg.

Preparation of Culture Media

About 14 g of nutrient agar was weighed and dispensed into 420 ml of water in a conical flask and was stirred using the glass rod, it was melted on the hotplate before autoclaving at 121 °C, in a bijoux bottle for an hour and was allowed to cool (Ponder, 2001).

Sensitivity Test

The method employed in the determination of antimicrobial activity using agar-well diffusion method is based on Basri, *et al.* [26]. 18 petri dishes were used for the growth of each bacterial species. 20ml of nutrient agar mixed with test organism was poured into sterilized petri dishes and was allowed to solidify. The wells of 6mm in diameter were bored into the agar media with sterilized cork borer. Each plate for each tested extract comprised five wells to accommodate the extract concentrations at 400 mg/ml, 200 mg/ml, 100 mg/ml and 50 mg/ml and the control. The plates were pre-incubated for 1 hour, allowing the complete diffusion of the samples before incubated at 37 °C overnight. The diameter of inhibition zone surrounding the well was measured to the nearest millimeter.

Determination of Minimum Inhibitory Concentration (MIC)

The minimum inhibitory concentration was determined to ascertain the lowest concentration of the plant extract that would allow growth or inhibition of each test organisms.

Double dilutions of the extracts starting from 400 mg/ml, 200 mg/ml, 100 mg/ml, 50 mg/ml, 25 mg/ml, 12.5 mg/ml and 6.25 mg/ml were prepared in a sterilized test tubes containing the sterilized nutrient broth. An accurate volume of 0.1 ml of the test organism was added to the respective set of test tubes.

After shaking, the test tubes were incubated at 37 °C for 24 hours. The test tubes were examined for turbidity. The presence of turbidity indicated growth on the test bacteria. The lowest concentration that prevented visible growth of the test organism was noted and taken as the minimum inhibitory concentration on that particular organism [27].

Determination of Minimum Bactericidal Concentration (MBC)

Tubes with no turbidity from minimum inhibitory concentration were taken for each organism. One loopful from each was taken and streaked on petri plates with nutrient agar, incubated for 24 hours at 37 °C. The plates were observed and any plate with growth of organism is said to be bacteriostatic at the concentration of the plant extract and any plate without growth signifies that the plant extract killed the bacteria at that concentration and is said to be bactericidal.

Results

The field survey revealed that the major occupation of respondents is farming, all from the same tribe (Afizere). The special name given to the plant is 'refat' and most varieties are found in the dry season majority of the respondents used the fruit as medicine, snack and obtained oil from it (Table 1). Most parts of the plant used are fruit, bark and leaf (Figure 1) [28]. Attributes of the plant identified showed that 80 % of the varieties are oily, 60 % sweet and 66.7 % hard (longer time of pre-warming) (Figure 2).

Plates 1 to 6 showed the fruits with different varieties (Figure 3).

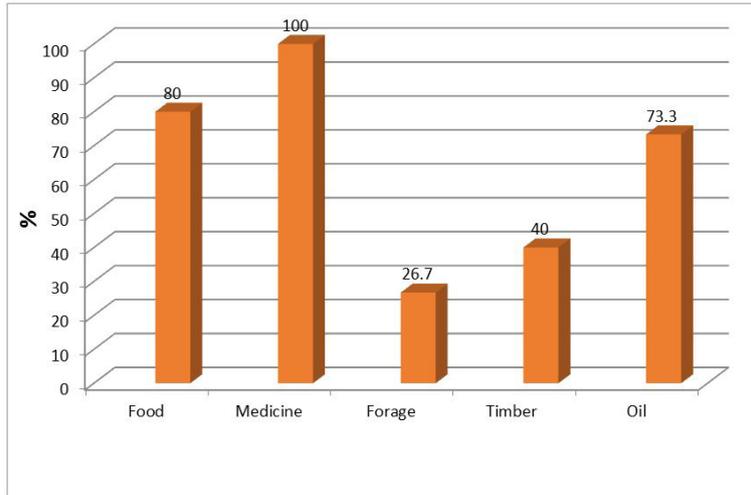


Figure 1: Uses of *Canarium schweinfurthii* in Fobur District of Jos East LGA of Plateau State

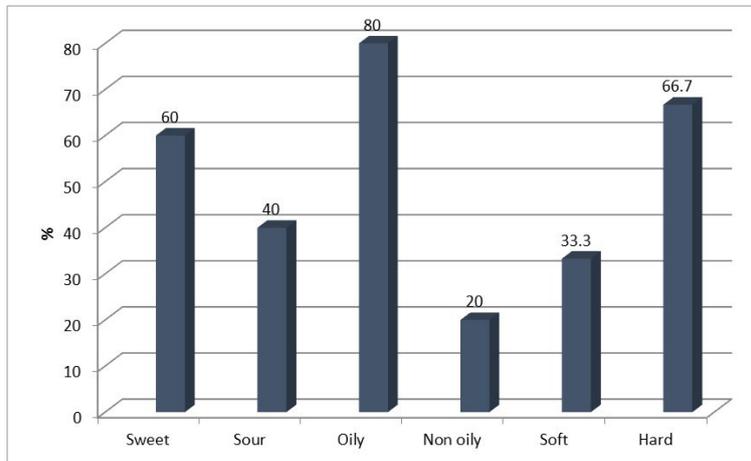


Figure 2: Attributes of *Canarium schweinfurthii* fruits



Figure 3: Collection of *Canarium schweinfurthii* fruit samples from different villages A&B. Rizek 1; Nubatong 2 (A&B); C. Kerker 3 (A&B)

Characteristics	Frequency	Percent
Age group		
15-24	3	20.0
25-34	3	20.0
35-44	5	33.3
45+	4	26.7
Education		
Primary	5	33.3
Secondary	6	40.0
Tertiary	4	26.7
Occupation		
Farming	9	60.0
Trading	2	13.3
Student	4	26.7
Tribe		
Afizere	15	100
Common name		
Atili	15	100
Special name		
Refat	15	100
When a Particular variety is found		
1 Dry season	13	86.7
2 Wet season	2	13.3
Uses of <i>Canariumschweinfurthii</i>		
Food	12	80.0
Medicine	15	100.0
Forage	4	26.7
Timber	6	40.0
Oil	11	73.3
Part use		
Leaf	7	46.7
Seed	1	6.7
Bark	14	93.3
Fruit	15	100.0
Harvesting		
November	4	26.7
December	7	46.7
January	9	60.0
February	4	26.7
Pest that attack ripe fruit		
Birds	7	46.7
Rodents	6	40.0
Others	6	40.0
Peak		
Wet season	2	13.3
Dry season	13	86.7
Attribute		
Sweet	9	60.0

Characteristics	Frequency	Percent
Sour	6	40.0
Oily	12	80.0
Non oily	3	20.0
Soft	5	33.3
Hard	10	66.7

Table 1: Result of background characteristic of Respondents

The ripening period of the varieties is between October to March Source: Field Survey, (2017)

Table 2 revealed the fruit characterization based on weight, width, length shapes and their respective pre-warming duration. The variety from Kerker had the highest weight 62 g followed by Nubatong (34.5 g) and Rizek (28 g). The shape of the fruit from Kerker and Rizek are ovate. While Nubatong is ellipse in shape.

Varieties	Weight (g)	Width (cm)	Shape	Duration/Prewarming at 18 °C
Kerker	6.22	7.5	Ovate	8 minutes
Nubatong	34.5	6.3	Elipse	12 minutes
Rizeck	28.5	6.1	Ovate	11 minutes

Table 2: Average fruit weight, length, width, shapes and duration of pre-warming

Source: Field Survey, 2017

Tables 3 and 4 showed the preliminary phytochemical screening of the plant extracts (leaf and fruit) of *Canarium schweinfurthii* showing the presence of flavonoids, tannins, saponins, carbohydrates, steroids, Anthraquinones and Terpenoids while alkaloids and cardiac glycosids were absent in leaf. While in fruits, phytochemicals revealed the presence of flavonoids, carbohydrates and steroids only.

Constituents	Location					
	Kal	Kbl	Nal	Nbl	Ral	Rbl
Alkaloids	-	-	-	-	-	-
Tannins	+++	+++	++	+++	++	++
Saponins	++	++	++	+	++	++
Flavonoids	++	+++	++	+++	++	+++
Carbohydrates	++	+++	+++	++	+++	++
Cardiac glycosides	-	-	-	-	-	-
Steriods	++	+++	++	+	++	+++
Anthraquinones	++	+	++	+	+++	++

Key

-	=	Absent
+	=	Present
++	=	Appreciably present
+++	=	Abundantly present
Kal	-	Leaf variety 1 from Kerker village
Kbl	-	Leaf variety 2 from Kerker village
Nal	-	Leaf variety 1 from Nubatong village
Nbl	-	Leaf variety 2 from Nubatong village
Ral	-	Leaf variety 1 from Rizek village
Rbl	-	Leaf variety 2 from Rizek village

Table 3: Result showing the phytochemical screening of the ethanolic leaf extracts of *Canarium schweinfurthii*

Constituents	Location					
	Kaf	Kbf	Naf	Nbf	Raf	Rbf
Alkaloids	-	-	-	-	-	-
Tannins	-	-	-	-	+	-
Saponins	-	-	-	-	-	-
Flavonoids	+	+	+	+	+	+
Carbohydrates	++	+	+++	+++	+++	++
Cardiac glycosides	-	-	-	-	-	-

Constituents	Location					
	Kaf	Kbf	Naf	Nbf	Raf	Rbf
Steriods	++	+	++	++	+	+
Anthraquinones	-	-	-	-	-	-
Terpenoids	-	-	-	-	-	-

Key
 - = Absent
 + = Present
 ++ = Appreciably present
 +++ = Abundantly present
 Kaf - Fruit variety 1 from Kerker village
 Kbf - Fruit variety 2 from Kerker village
 Naf - Fruit variety 1 from Nubatong village
 Nbf - Fruit variety 2 from Nubatong village
 Raf - Fruit variety 1 from Rizek village
 Rbf - Fruit variety 2 from Rizek village

Table 4: The result showing the phytochemical screening of the fruits extracts of *Canarium schweinfurthii*

The Table 4 revealed the effect of extracts on test organism. The leaf extract from Kerker is not sensitive on *Streptococcus mutans* at all concentrations. All organisms showed sensitivity at all concentrations except *Escherichia coli* at 100 mg/ml and 50 mg/ml. The controls inhibit all test organisms.

The Table 5 above showed the sensitivity of leaf extract from Nubatong village (Table 6). The extract showed inhibition on *Streptococcus mutans*, *Bacillus subtilis*, and *Escherichia coli*. There was no inhibition on *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Salmonella typhi*. All organisms were inhibited by the control (Plate 8) (Figure 4).

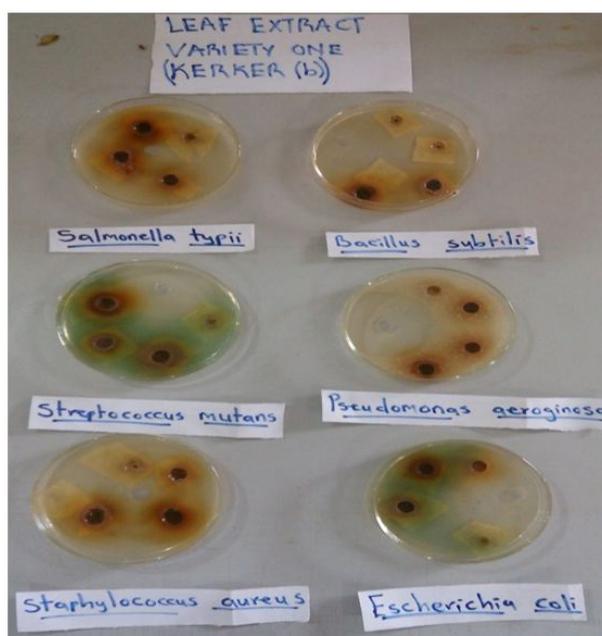


Figure 4: Plate 8- Zone of inhibition of *Canarium schweinfurthii* leaf extracts from Kerker village and the control

Test Organism	Mean diameter of the zones of inhibition (mm) at different concentration (mg/ml) of extract				Control Ciprofloxacin
	400	200	100	50	
<i>Pseudomonasaeruginosa</i>	22	20	15	13	50
<i>Staphyloccusaureus</i>	20	23	14	10	32
<i>Streptococcus mutans</i>	-	-	-	-	50
<i>Salmonella typhi</i>	29	26	21	14	30
<i>Bacillus subtilis</i>	27	22	18	12	34
<i>Escherichia coli</i>	25	20	-	-	52

Table 5: Antibacterial activity of Ethanolic leaf extract of *Canarium schweinfurthii* on Kerker village variety on test organisms

Test Organism	Mean diameter of the zones of inhibition (mm) at different concentration (mg/ml) of extract				Control Ciprofloxacin
	400	200	100	90	
<i>Pseudomonasaeruginosa</i>	-	-	-	-	51
<i>Staphylococcus aureus</i>	-	-	-	-	23
<i>Streptococcus mutans</i>	18	14	10	8	63
<i>Salmonella typhi</i>	-	-	-	-	35
<i>Bacillus subtilis</i>	12	10	7	5	24
<i>Escherichia coli</i>	28	24	20	15	37

Table 6: Antibacterial activity of Ethanolic leaf extract of *Canarium schweinfurthii* on Nubatong village variety on test organism

Table 7 indicated the antibacterial activity of extract from Rizek village. *Pseudomonasaeruginosa* was not inhibited by the extract at all concentration including the control. All test organisms were inhibited at all concentration but *Bacillus subtilis* was not inhibited at 100 mg/ml, and 50 mg/ml (Plate 7) (Figure 5).

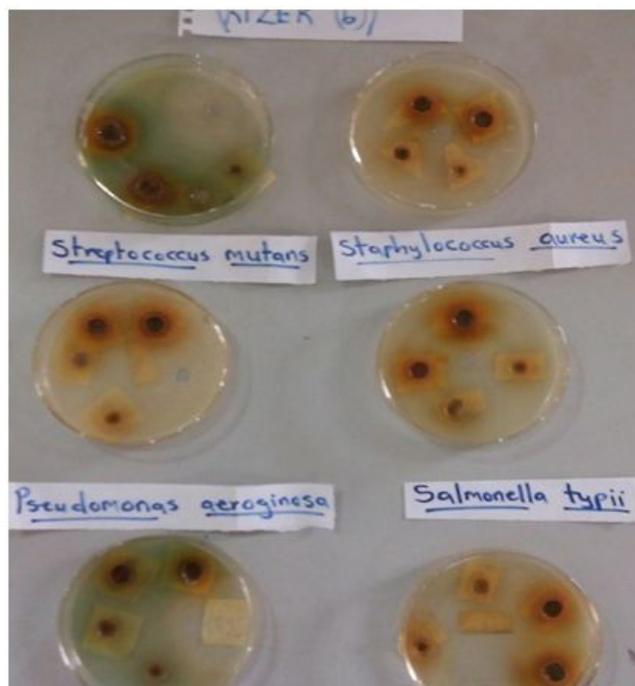


Figure 5: Plate 7- Zone of inhibition of *Canarium schweinfurthii* leaf extracts from Rizek village and the control

Test Organism	Mean diameter of the zones of inhibition (mm) at different concentration (mg/ml) of extract				Control Ciprofloxacin
	400	200	100	90	
<i>Pseudomonasaeruginosa</i>	-	-	-	-	-
<i>Staphylococcus aureus</i>	40	31	28	23	45
<i>Streptococcus mutans</i>	32	31	28	23	55
<i>Salmonella typhi</i>	28	27	25	15	30
<i>Bacillus subtilis</i>	20	15	-	-	35
<i>Escherichia coli</i>	15	10	9	7	38

Table 7: Antibacterial activity of Ethanolic leaf extract of *Canarium schweinfurthii* (Rizek variety) on test organism

The Table 7 above indicated the minimum inhibitory concentration of test organism on leaf extract from Kerker. There was no growth of test organism between 400mg/ml and 50mg/ml. At 25 mg/ml *Pseudomonas aeruginosa*, *Salmonella typhi* and *Bacillus subtilis* showed minimum inhibitory at 50 mg/ml. While *Staphylococcus aureus* showed (MIC) at 25 mg/ml. *Streptococcus mutans* and *Escherichia coli* showed MIC at 12.5 mg (Figure 6).

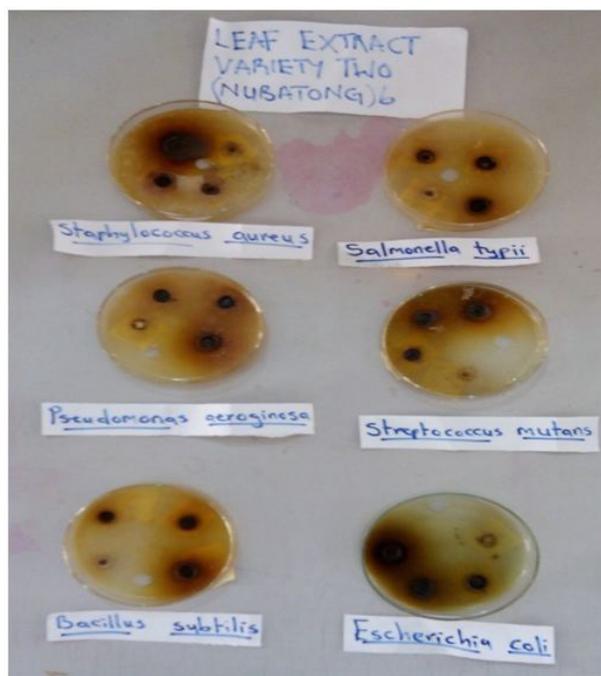


Figure 6: Plate 9- Zone of inhibition of *Canarium schweinfurthii* leaf extracts from Nubatong village and the control

The Table 8 showed Minimum Inhibitory Concentration (MIC) on *Streptococcus mutans* and *Escherichia coli* at 12 mg/ml, *Bacillus subtilis* at 50 mg/ml.

Test Organism	Concentration on (mg/ml)							MIC
	400	200	100	50	25	12.5	6.25	
<i>Pseudomonasaeruginosa</i>	-	-	-	-	+	+	+	50
<i>Staphylococcus aureus</i>	-	-	-	-	-	+	+	25
<i>Streptococcus mutans</i>	-	-	-	-	-	-	+	12.5
<i>Salmonella typhi</i>	-	-	-	-	+	+	+	50
<i>Bacillus subtilis</i>	-	-	-	-	+	+	+	50
<i>Escherichia coli</i>	-	-	-	-	-	-	-	12.5

Key
 + = Growth (inhibition)
 - = No Growth

Table 8: Minimum Inhibitory Concentration (MIC) of the ethanolic leaf extract of *Canarium schweinfurthii* on Kerker village variety

The Table 9 showed the MIC of *Staphylococcus aureus* at 100 mg/ml followed by *Escherichia coli* 50 mg/ml. *Bacillus subtilis* and *Streptococcus mutans* at 25 mg/ml while *Salmonella typhi* showed MIC at 12.5 mg/ml.

Test Organism	Concentration on (mg/ml)							MIC
	400	200	100	50	25	12.5	6.25	
<i>Streptococcus mutans</i>	-	-	-	-	-	+	+	2.5
<i>Bacillus subtilis</i>	-	-	-	-	+	+	+	50
<i>Escherichia coli</i>	-	-	-	-	-	-	+	12.5

Key
 + = Growth (inhibition)
 - = No Growth

Table 9: Minimum Inhibitory Concentration (MIC) of the ethanolic leaf extract of *Canarium schweinfurthii* on Nubatong village variety

The Minimum Bactericidal Concentration (MBC) on leaf extract from Kerker village showed that *Pseudomonas aeruginosa*, *Salmonella typhi* and *Bacillus subtilis* had 25 mg/ml while *Staphylococcus aureus* and *Escherichia coli* had 12.5 mg/ml respectively.

The result showed the Minimum Bactericidal Concentration of *Streptococcus mutans* and *Escherichia coli* at 12.5 mg/ml and *Bacillus subtilis* at 25 mg/ml.

Test Organism	Concentration on (mg/ml)							MIC
	400	200	100	50	25	12.5	6.25	
<i>Staphylococcus aureus</i>	-	-	-	+	+	+	+	100
<i>Streptococcus mutans</i>	-	-	-	-	-	+	+	25
<i>Salmonella typhi</i>	-	-	-	-	-	-	+	12.5
<i>Bacillus subtilis</i>	-	-	-	-	-	+	+	25
<i>Escherichia coli</i>	-	-	-	-	+	+	+	50

Key

+ = Growth (inhibition)

- = no growth

Table 10: Minimum Inhibitory Concentration (MIC) of the ethanolic leaf extract of *Canarium schweinfurthii* on Kerker variety

Test Organism	Concentration on (mg/ml)							MBC
	400	200	100	50	25	12.5	6.25	
<i>Pseudomonasaeruginosa</i>	-	-	-	-	-	+	+	25
<i>Staphylococcus aureus</i>	-	-	-	-	-	-	+	12.5
<i>Salmonella typhi</i>	-	-	-	-	-	+	+	25
<i>Bacillus subtilis</i>	-	-	-	-	-	+	+	25
<i>Escherichia coli</i>	-	-	-	-	-	-	+	12.5

+ = Growth (inhibition)

- = No Growth

Table 11: Minimum Bactericidal Concentration (MBC) of the ethanolic leaf extract of *Canarium schweinfurthii* on Kerker village variety Key

The Table 12 showed Minimum Bactericidal Concentration of *Pseudomonas aureginosa* and *Salmonella typhi* at 25 mg/mg. The MBC of *Staphylococcus aureus*, *Bacillus subtilis* and *Escherichia coli* at 12 mg/ml (Table 13).

Test Organism	Concentration on (mg/ml)							MIC
	400	200	100	50	25	12.5	6.25	
<i>Streptococcus mutans</i>	-	-	-	-	-	-	+	12.5
<i>Bacillus subtilis</i>	-	-	-	-	-	+	+	25
<i>Escherichia coli</i>	-	-	-	-	-	-	+	12.5

+ = Growth (inhibition)

- = No Growth

Table 12: Minimum Bactericidal Concentration (MBC) of the leaf ethanolic extract of *Canarium schweinfurthii* on Nubatong village variety

Test Organism	Concentration on (mg/ml)							MIC
	400	200	100	50	25	12.5	6.25	
<i>Pseudomonasaeruginosa</i>	-	-	-	-	-	+	+	25
<i>Staphylococcus aureus</i>	-	-	-	-	-	-	+	12.5
<i>Salmonella typhi</i>	-	-	-	-	-	+	+	25
<i>Bacillus subtilis</i>	-	-	-	-	-	-	+	12.5
<i>Escherichia coli</i>	-	-	-	-	-	-	+	12.5

Key

+ = Growth (inhibition)

- = No Growth

Table 13: Minimum Bactericidal Concentration (MBC) of the leaf ethanolic extract of *Canarium schweinfurthii* on Rizek village variety

Discussion

With respect to the local people's knowledge on *Canarium schweinfurthii*, the study showed that all respondents affirmed that the plant can be used for medicine, food, oil, timber and forage. It also revealed that the part of plant mostly used are the fruit and the bark, these findings have also been reported by Dawang, *et al.* [11]. The special names given to this plant 'refat' have been previously reported by Nyam, *et al.* [29]. The study also revealed that the different types of varieties based on shape, weight, length, width of the fruit, this is also supported by Maduelosi and Angaye [9]. The duration of prewarming on the varieties revealed between 8 to 11 minutes at 18 -20 °C which has also been affirmed by Nyam, *et al.* [8].

The phytochemical screening of results on leaves of ethanolic extracts showed the presence of chemical active compounds such as Tannins, Saponins, flavonoids, carbohydrates, steroids and anthraquinones [13,30]. Alkaloid and Cardiac Glycosides were not detected in the leaf extract as also reported by Koto-te-Nyiwa, *et al.* [31]. The phytochemical screening on fruits in ethanolic extracts only showed the presence of carbohydrates, flavonoids and steroids other chemical compounds were absent. The result of phytochemicals had also been reported to be responsible for the growth inhibition of numerous micro-organisms in their cell membrane [32,33].

The test organisms used were all susceptible to the leaf extracts of *Canarium schweinfurthii*, among the extract, the variety from Rizek village exhibited highest zone of inhibition with 40mm against *Staphylococcus aureus* followed by variety from Kerker village and variety from Nubatong exhibiting no zone of inhibition against *staphylococcus aureus*. All the test organisms except *Pseudomonas aeruginosa* were susceptible to the leaf extract and this could be attributed to the differences in their cell wall composition as affirmed by Koko-te-Nyiwa, *et al.* [31].

The Minimum Inhibitory Concentration (MIC) results revealed that the ethanolic extract of the leaves of *Canarium schweinfurthii* inhibited the test organisms at a high concentration of 40mg/ml and low concentration between 25 mg/ml to 6.25 mg/ml showed no susceptibility. Morgan & Wiart confirmed the potency of the leaf against Gram positive bacteria *Staphylococcus aureus* and gram negative bacteria *Pseudomonas aeruginosa* on ethanol extracts [34,35].

Minimum bactericidal concentration of 25 mg.ml and 12.5 mg/ml showed bactericidal activity against all test organisms has also reported by Den, *et al.* and Okoli, *et al.* [36,37].

Conclusion

This study has revealed pool of information on the different variety of *Canarium schweinfurthii* found in the study area. It also revealed its antimicrobial tendency with increased ethanolic leaf concentration against some microorganisms.

Recommendations

Based on the findings from the study, further research should be carried out on the identification and documentation of the leaf varieties. Sensitivity test should be carried out using the fruits as well as the toxicity of other parts together with other methods of extraction should be carried out. Other clinical microbial species should also be studied using the different plant parts. Also *Canarium schweinfurthii* should be planted on a large scale to ensure its sustainable utilization.

References

- Kochar SL (1981) Tropical Crops: A Textbook of Economics Botany/ International College Editions, Macmillan, London, UK.
- Sofowora AO (1993) Medicinal Plants and Traditional Medicine in Africa. Spectrum Books Ltd., Ibadan, Nigeria. 191-289.
- Choudhary SE, Singh RN, Upadhyang RK, Singh RK, Choudhang HR, et al. (2014) Effect of vegetable. Intercrops and Planting Pattern of maize on Growth Yield and Economics of water maize in Eastern Uttar Pradesh. Environment and Ecology 32: 101-5.
- Keay RWJ (1989) Trees of Nigeria. A revised version of the Nigerian trees. Clarendon Press Oxford 476-8.
- Orwa CA, Mutua K, Jammadass R, Anthony RS (2009) Agroforestry Data base. A Tree reference and selection guide version 4.0. Retrieved 8th of December 2016.
- Nyam MA (2011) This Effects of Microbial Colonization of *Canarium schweinfurthii* Linn ('Atili') Fruit oil on its Domestic and Industrial Uses. Ph.D Thesis, University of Jos, Nigeria.
- Burkill HM (1985) The useful plants of West Tropical Africa. (2nd edn) Vol1 families A – D Royal Botanic Gardens, Kew, Richmond, United Kingdom 96.
- Nyam MA, Makut MD, Itelima JU, Daniel AM (2014) Nutritional potential of the fruits of Black olive (*Canariumschweinthurfii* Linn) from Plateau State, Nigeria. Pakistan J Nutri 13: 335-9.
- Maduelosi NJ, Angage SS (2015) Characterization of African Elemi *Canariumschweinfurthii*. International J Adv Res Chem Sci 2: 34-6.
- Nyam MA, Wonang DL (2004) Proximate Chemical Analysis of ('Atili') *Canariumschweinfurthii* Linn. Endosperm and Elemental Analysis of the Oil. Nutrition Society of Nigeria 35: 78-81 Proceeding of 35yh Annual Conference.
- Dawang SN, Danahap TS, Makrerang SS, Nyam MA (2016) Preliminary survey of the indigenous knowledge of *Canariumschweinfurthii* (Engl) in some parts of Plateau State, Nigeria. IOSR J Pharm BiolSci 11: 76-82.
- Morikawa M (2006) Beneficial Biofilm formation by Industrial Bacteria *Bacillus subtilis* and Related species. J Biosci Bioeng 1: 1-8.
- Stockwell VO, Sugar D, Spotts R, Johnson KB, Loper JE (1998) Recovery of streptomycin – resistant isolates of *Erwiniaamylovora* from Oregon orchards. Phytopathology 80: 550.
- Moreman DE (1996) An analysis of the Flood Plants and Drug Plants of Native North America. J Ethnopharmacol 52: 1-22.
- Newman DJ, Craig GM, Snader KM (2003) Natural Products as sources of new Drug over Period 1981-2002. J Natural Product70: 660-037.
- Martin FN, Campbell CW, Ruberte RM (1987) Perennial edible fruits of the tropics: An inventory. USDA. Agric. Handbook 642- ARS, Washington DC, USA.
- George D, Pumploma-Roger MD (1999) Encyclopedia of Medicinal Plants. Education and Health library Spain 2: 600-45.
- Igoli JO, Tor-Anyin TA, Usman SS, Oluma HO A, Igoli NP (2002) Folk medicine of the lower Benue valley of Nigeria. In Recent progress in Medicine Plants Vol. 1 Ethnomedicine and Pharmacognosy Part II 371-38.
- Lewis WH, Elvin-Lewis MP (1999) Medicinal Plants as Sources of New Therapeutics. Ann MO Bot Grad 83: 16-24.
- Bandow JE, Brötz H, Hecker M (2002) *Bacillus subtilis* Tolerance of Moderate concentrations of Rifampin Involves the B-Depent general and Multiple Stress Response. J Bacteriol 184: 459-67.

21. Abayeh OJ, Abdulrazaq AK, Olaogun R (1999) Quantity Characteristics of CanariumschweinfurthiiEngl oil. *Plant Foods Hum Nutr* 54: 43-7.
22. Johnson G, Olsen R (1997) Multiple pathways for Toluene Degradation in Burkholderia sp. Strain IS 150 *Applied and Environmental Microbiology* 268-72.
23. *Journal of the American oil Chemists' Society*.www.springer.com>chemistry>journal> (18/04/2017).
24. Harbone JE (1983) *Phytochemicals Methods* Second edition, Academic Press, London.
25. Trease GE, Evans NC (1989) *Pharmacognosy*, (11th edn) Bailliera Tindall, London 45-50.
26. Basri DE, Saidi N, Mahari H, Saari S, Santhanam J (2014) Preliminary screening for Antimicrobial Activity of the Pulp of CanariumodontophylumMiq (Dabai) Fruit. *Global J Pharmo* 8: 213-20.
27. Ehiem JC, Ndirika IO, Onwuka UN (2016) Effect of moisture content on some physical properties of Canarium schweinfurthii Engl Fruit *Res Agronomy Eng* 62: 162-9.
28. Monica C (2000) *Antibacterial sensitivity testing. District Laboratory Practice in tropical Countries Part 2: Cambridge University Press. Cape town, South Africa.* 105-97.
29. Nyam MA, Wonang DL, Akueshi CO (2009) Phytochemical screening and Antimicrobial Studies on Canariumschweinfurthii.('Atili') fruits and oil. *Nig J Botany* 22: 247-53.
30. Wall ME, CR, McClema ML, Klong ME (1992) Detection and estimation of steroid and saponins in plant tissue. *Anal Chem* 24: 1337-42.
31. Koto-te-Nyiwa N, Lengbiye EM, Joseph KL, Pius TM (2015) Canarium schweinfurthii Engl. (Burseracias): An updated Review and future Direction of Sickle Cell Disease. *J Adv Med Life Sci*.
32. Ibara JR, Alion-Hou RDG, Ouamba JM, Diatwa M, Gbeassor M, et al. (2007) Preliminary Evaluation of Antulcerogenic Activity of CeibaPentandra. *J Med Sci* 7: 485-8.
33. Kunle OF, Egbarevba HO (2009) *Ethnobotanical leaflets; Department of Medical Plant Research and Traditional, National Institute of Pharmaceutical Research and Development, Nigeria* 13: 1216-21.
34. Mogan R, Wiart C (2011) Canarium L.A phytochemical and Pharmacological Review. *J Pharm Res* 4: 2482-9.
35. Lederberg J (2000) *Pseudomonas. Encyclopedia of Microbiology Second edition. Vol-3 San-Diego Pp* 816-91.
36. Deng W, Liou SR, Plunkett G 3rd, Mayhew GF, Rose DJ, et al. (2003).Comparative Genomics of Salmonella enteric Serovar. Typhi strains Ty2 and CT 18. *J Bacteriol*185: 2330-7.
37. Okoli BJ, Ayo RG, Habila JD, Ndukwe GI, Jummai AT (2015) Inhibition of Methicithin Resistance Staphylococcus aureus and Fungi by Cannarium schweinfurthii extracts. *Scholar Acad J Biosci Sch Acad J Bioscie* 3: 413-25.